

Review

Prognostic molecular markers in early breast cancer

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Published: 11 March 2004

Breast Cancer Res 2004, **6**:109-118 (DOI 10.1186/bcr777)

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Abstract

A multitude of molecules involved in breast cancer biology have been studied as potential prognostic markers. In the present review we discuss the role of established molecular markers, as well as potential applications of emerging new technologies. Those molecules used routinely to make treatment decisions in patients with early-stage breast cancer include markers of proliferation (e.g. Ki-67), hormone receptors, and the human epidermal growth factor receptor 2. Tumor markers shown to have prognostic value but not used routinely include cyclin D₁ and cyclin E, urokinase-like plasminogen activator/plasminogen activator inhibitor, and cathepsin D. The level of evidence for other molecular markers is lower, in part because most studies were retrospective and not adequately powered, making their findings unsuitable for choosing treatments for individual patients. Gene microarrays have been successfully used to classify breast cancers into subtypes with specific gene expression profiles and to evaluate prognosis. RT-PCR has also been used to evaluate expression of multiple genes in archival tissue. Proteomics technologies are in development.

Keywords: biological tumor markers, breast cancer, genomics, prognosis, proteomics

Introduction

Breast cancer is the most common malignancy in women, and it is highly curable if diagnosed at an early stage. Traditional prognostic factors include the axillary lymph node status, the tumor size, and the nuclear grade and histologic grade. Interest in novel prognostic markers is based on the fact that a significant number of patients with early-stage breast cancer harbor microscopic metastasis at the time of diagnosis. It is now well established that adjuvant systemic therapy improves survival in patients with early-stage breast cancer [1,2]. Treatment options for early-stage breast cancer include chemotherapy (e.g. anthracyclines, taxanes) and hormone therapy (e.g. tamoxifen, aromatase inhibitors). The use of trastuzumab is under investigation in the adjuvant setting for patients with human epidermal growth factor receptor (HER) 2 overexpressing breast cancer.

Systemic therapies are potentially toxic, however, and identifying the individual patients who are at high risk and likely to benefit remains a major challenge. For example, the risk of recurrence for a patient with negative axillary lymph nodes and a tumor measuring 1–2 cm is approximately 20–30%. Most patients in this group are currently offered adjuvant systemic therapy, although up to 70% of patients would not need it because they are already cured of their disease. Unfortunately, the histologic information is clearly not sufficient to accurately assess individual risk and to possibly avoid adjuvant systemic therapy. A large number of molecular markers have been studied to determine their ability to predict prognosis or response to therapy, or both (Table 1). Prognostic factors correlate with survival independent of systemic therapy, and are used to select patients at risk. Predictive factors correlate response to therapy independent of prognosis, and have a

DFS = disease-free survival; ELISA = enzyme-linked immunosorbent assay; ER = estrogen receptor; HER = human epidermal growth factor receptor; IHC = immunohistochemistry; PAI = plasminogen activator inhibitor; PCR = polymerase chain reaction; PR = progesterone receptor; RT = reverse transcription; uPA = urokinase-type plasminogen activator.

Table 1**Well-established and investigational prognostic factors in breast cancer**

Well-established prognostic factor	Investigational prognostic factor
Ki-67	pS2
Estrogen receptor	Mitotin
Progesterone receptor	Epidermal growth factor receptor
HER-2	Insulin-like growth factors
	Apoptosis-related proteins
	Cell cycle molecules
	Plasminogen activators and inhibitors
	Angiogenesis-related proteins

significant impact in selected patient populations. Some molecular markers are associated with prognosis, some are associated with response to therapy, and some are associated with both.

Although a large number of molecules have been investigated as potential prognostic and predictive factors, the National Institute of Health Consensus Development Conference held in 2000 stressed the need for validation and appropriate quality control for most of the markers studied to date [3]. The present article reviews the available data on established and investigational prognostic molecular markers in patients with early-stage breast cancer.

Proliferation markers

The tumor proliferation rate is an important prognostic factor in breast cancer. Several methods have been developed to estimate the proliferative rate of tumor cells. The S-phase fraction, as measured by flow cytometry, is a validated method for measuring tumor proliferation [4]. However, flow cytometry is not commonly used because of the amount of tissue consumed for the assay. Alternative methods for measuring tumor proliferation have been developed, including immunohistochemistry (IHC) to detect cell cycle-related antigens, that are better suited for the evaluation of small archival tissue samples.

Ki-67 is a nuclear antigen found in cells in the proliferative phases of the cell cycle (G1 phase, S phase, G2 phase, and M phase) but not cells in the resting phase (G0 phase). MIB-1 is a monoclonal antibody that identifies the Ki-67 protein in paraffin-embedded tissue. A strong correlation has been noted between the percentage of cells showing Ki-67 staining and the nuclear grade, age, and mitotic rate [5,6]. Patients whose tumors overexpress Ki-67 in more than 50% of the cells are at high risk of developing recurrent disease [7]. In addition, Ki-67

correlates with other well-characterized proliferation markers, such as the proliferating cell nuclear antigen [6].

Mitotin, a recently described 350-kDa nuclear phosphoprotein, is expressed in the late G1 phase, S phase, G2 phase, and M phase of the cell cycle, but not in the G0 phase [8]. Clark and colleagues [9] showed that mitotin is a proliferation marker that correlates with high S-phase fraction and negative estrogen receptor (ER)/progesterone receptor (PR) status. Although mitotin was not a predictor of survival in the study by Clark and colleagues, it was an independent predictor of recurrence. Additional studies are necessary to validate these findings.

Estrogen receptors and progesterone receptors

Estrogen mediates its functions through two specific intracellular receptors, the ER α and the ER β , which act as hormone-dependent transcriptional regulators [10,11]. The ER pathway plays a critical role in the pathophysiology of human breast cancer. Overexpression of ER α is a well-established prognostic and predictive factor in breast cancer patients. The prognostic significance of ER β is not well defined [12–15]. Overexpression of the PR serves as a functional assay because it indicates that the ER pathway is intact, even if the tumor is reported as ER-negative. When biochemical ligand-binding assays indicate concentrations of 10 fmol/mg cytosol protein or more, the tumors are generally considered ER-positive and PR-positive for clinical purposes.

The ER and PR status can be measured using IHC. The results of IHC correlate closely with biochemical ligand-binding assays and with clinical response rates to endocrine therapy [16]. Because IHC, unlike chemical assays, does not require the destruction of tissue specimens, and because it shows the tissue distribution of ER, it has become the preferred method for determining the ER/PR status in breast cancer specimens. Quantitative methods using computer-aided image analysis are being developed to improve the accuracy of IHC.

The value of ER status as an independent prognostic variable is diminished by its association with other established indicators of favorable prognosis. These include older age, low-grade histology, a favorable nuclear grade, a low S-phase fraction, a normal complement of DNA, a low proliferative index, and a low thymidine-labeling index [17]. In addition, ER-positive patients receive and benefit from either adjuvant or palliative hormone therapy so regularly that it is difficult to evaluate the prognosis apart from the influence of therapy.

In some studies, the higher disease-free survival (DFS) and overall survival rates of patients with ER-positive tumors are seen only in the presence of hormone therapy. Often the favorable effect of ER-positive status as a

discriminant is lost after several years, suggesting that the influence of treatment is temporary [18,19]. When node-positive patients not receiving adjuvant hormone therapy were studied, the 5-year DFS rate was 20% higher for ER-positive patients compared with that for ER-negative patients. However, the 5-year DFS rate of the most favorable subgroup (i.e. patients with one to three positive nodes and ER-positive tumors) did not exceed 60% [20].

Among node-negative patients, small but statistically significant differences in DFS and overall survival rates have been found between ER-positive cases and ER-negative cases after various periods of follow-up [21]. The results of a multivariate analysis of prognostic factors by McGuire and colleagues [22], including the ER status for more than 3000 patients, showed the ER status to be more important for prognosis than tumor size in node-negative cases but not in node-positive cases. In one study, the ER status was found to be less important for predicting duration of DFS or overall survival than the nuclear grade and the number of positive nodes [23]. Allred and colleagues [24] showed that tamoxifen decreased the risk of local-regional recurrence in patients with ER-positive ductal carcinoma *in situ*.

The ER IHC assay is not standardized. Methods of tissue procurement, preservation, antigen retrieval, and, more importantly, the definition of positivity vary between different laboratories.

The prognostic role of ER β is not well defined. Fuqua and colleagues [25] evaluated ER β expression using IHC in 242 breast cancer patients. Their study showed that most tumors coexpressed both ER α and ER β . Although ER α expression was positively correlated with low tumor grade, with diploidy, and with low S-phase fraction (all biological parameters of a good prognostic profile), ER β trended toward an association only with aneuploidy. No association with tumor grade or S-phase fraction was seen for ER β . Larger studies are needed to determine the clinical utility of ER β expression in breast cancer.

Human epidermal growth factor receptor 2

The most frequently implicated receptors and growth factors in human breast cancer are members of the epidermal growth factor receptor subfamily of tyrosine kinase receptors. In addition to epidermal growth factor receptor, the type I subfamily includes HER-2, HER-3, and HER-4 [26,27]. These receptors share a common molecular architecture; they all possess a large glycosylated extracellular ligand-binding domain, a single hydrophobic transmembrane domain, and a cytoplasmic tyrosine kinase domain.

HER-2 (also known as *c-erbB-2* or *neu*) is a proto-oncogene that encodes a 185-kDa tyrosine kinase glycoprotein.

Amplification of the *HER-2* gene plays an important role in the pathogenesis of breast cancer [28–30]. The HER-2 protein is overexpressed in 60% of ductal carcinomas *in situ* and in 20–30% of infiltrating breast carcinomas [31,32]. The HER-2 status can be determined in human tumor samples using IHC or fluorescence *in situ* hybridization [32]. Amplification and/or overexpression of the HER-2 oncogene is associated with a poor DFS rate in patients with axillary node-positive breast cancer [32–34].

Allred and colleagues [35] evaluated HER-2 expression using IHC in 613 patients with node-negative breast cancer enrolled in the Intergroup Study 0011. In their study, patients were stratified into low-risk groups ($n=307$) and high-risk groups ($n=306$) on the basis of tumor size and ER status. Low-risk patients were defined as having small (<3 cm), ER-positive tumors and were observed without additional treatment after initial surgery. High-risk patients had either ER-negative tumors or large (≥ 3 cm), ER-positive tumors and were randomized to be observed ($n=146$) or to receive adjuvant chemotherapy ($n=160$) after surgery. In Allred and colleagues' study, HER-2 was overexpressed in 14.3% of all tumors combined, and overexpression was higher in invasive carcinomas associated with an extensive *in situ* component (21.5%) than in carcinomas without a significant noninvasive or *in situ* histologic component (11.2%; $P < 0.0001$). When patients with low-risk lesions not containing a significant *in situ* component ($n=179$) were analyzed, HER-2 was a strong prognostic factor. Patients in this group with HER-2-positive tumors showed only 40% DFS at 5 years, compared with more than 80% in patients with HER-2-negative tumors ($P < 0.0001$).

HER-2 overexpression has been associated with improved response to doxorubicin-based chemotherapy [36–40]. HER-2 overexpression does not seem to predict response to taxane-based chemotherapy [41]. The association between HER-2 overexpression and response to hormonal therapy is controversial [40,42,43]. Osborne and colleagues [44] reported an association between the ER coactivator AIB1 (SRC-3) and tamoxifen resistance, particularly in patients with HER-2-positive tumors treated with tamoxifen.

One of the main reasons for the clinical utility of the tissue measurement of HER-2 is for selection of patients with invasive breast cancer for trastuzumab monoclonal antibody therapy (Herceptin™; Genentech, San Francisco, CA, USA). The pivotal clinical trials of trastuzumab were conducted in patients with metastatic breast cancer overexpressing HER-2. The HER-2 status was determined by IHC using two monoclonal antibodies: CB-11 and 4D5. The HER-2 expression was scored as 0, 1+, 2+, and 3+, depending on the number of cells with membrane staining and on the intensity of the staining. If the tumor

was 0 or 1+, it was considered HER-2-negative. If the tumor was 2+ or 3+ it was considered HER-2-positive, and patients received trastuzumab therapy. Retrospective studies showed that the response rate for trastuzumab therapy was higher among patients with an IHC score of 3+ expression compared with patients having a score of 2+. Good correlation was noted between a 3+ score using IHC and the presence of HER-2 gene amplification using fluorescence *in situ* hybridization. Recent data indicate that fluorescence *in situ* hybridization may be a better predictor of response to trastuzumab-based therapy [45,46]. Trastuzumab is currently approved for treating patients with metastatic breast cancer. Adjuvant trials are ongoing to determine the safety and efficacy of trastuzumab in patients with early-stage breast cancer.

Plasminogen activators and inhibitors

Tumor cell invasion and metastasis is a multifactorial process that at each step may require the action of proteolytic enzymes, such as collagenases, cathepsins, plasmin, or plasminogen activators. Some of these molecules have been associated with specific prognoses and are now discussed in more detail.

The urokinase-type plasminogen activator (uPA) is a serine protease that plays an important role in the invasion and metastasis process through degradation of the extracellular matrix. High levels of tissue uPA and its inhibitors (plasminogen activator inhibitor [PAI]-1, PAI-2) measured using ELISAs have been correlated with poor outcome in node-negative breast cancer patients [47–51].

Janicke and colleagues [52] conducted a prospective, randomized multicenter clinical trial of adjuvant therapy versus observation for patients with node-negative breast cancer. In their study, patients whose primary tumors had low tumor levels of uPA and PAI-1 (low risk) did not receive adjuvant systemic therapy. Patients with elevated tumor levels of uPA and/or of PAI-1 (high risk) were randomized to receive cyclophosphamide, methotrexate, and 5-fluorouracil adjuvant chemotherapy or to receive no treatment. The first interim analysis showed an estimated 3-year recurrence rate of 6.7% in the low-risk group and of 14.7% in the high-risk group ($P=0.006$). The intent-to-treat 3-year DFS rate for patients in the high-risk group assigned to chemotherapy or to observation was not statistically different. When the results were analyzed based on actual treatment delivered, however, the 3-year DFS rate for patients treated with adjuvant chemotherapy was 9% versus 19% for patients who did not receive chemotherapy ($P=0.016$). The improvement in actuarial 3-year DFS was maintained at a median follow-up of 50 months [53].

Zemzoum and colleagues [54] showed that uPA/PAI-1 levels in primary tumor tissue are associated with an

Table 2

Pro-angiogenic and anti-angiogenic proteins

Pro-angiogenic protein	Anti-angiogenic protein
Vascular endothelial growth factor	Angiostatin
Angiogenin	Endostatin
Angiopoietin-1	Interferon (alpha, beta)
Del-1	Interleukin-12
Fibroblast growth factors	2-Methoxyestradiol
Follistatin	Platelet factor 4
Interleukin-8	Thrombospondin
Leptin	CD59 complement fragment
Placental growth factor	Heparinases
Platelet-derived endothelial growth factor	Tissue inhibitors of metalloproteinases
Pleiotrophin	Vasostatin
Transforming growth factor alpha	16-kDa prolactin fragment
Transforming growth factor beta	
Tumor necrosis factor	
Vascular endothelial growth factor	
Hepatocyte growth factor	
Nitric oxide	
Erucamide	
Urokinase plasminogen activator	

aggressive course of disease in lymph node-negative breast cancer, independent of HER-2 status. It has been suggested that patients with node-negative breast cancer and low levels of uPA and PAI-1 may be spared the trauma of adjuvant chemotherapy [55]. However, the ELISAs of uPA/PAI-1 require extracts of primary tumor tissue, and this is a major limitation for patients with small tumors.

Angiogenesis-related prognostic markers

It is now accepted that solid tumors must develop a vascular network to grow beyond 1 cm³, and they do so by stimulating the formation of new blood vessels (so-called angiogenesis). Angiogenesis is an active process, regulated by a large number of pro-angiogenic and anti-angiogenic molecules (Table 2). Interest in neovascularity as a prognostic factor was stimulated by the work of Folkman on tumor angiogenesis and by the potential for treatment with anti-angiogenic agents [56].

The prognostic relevance of tumor angiogenesis in breast cancer was first reported by Weidner and colleagues [57], who counted microvessels (veins and arteries) in the most densely vascularized areas of 49 invasive carcinomas and found their number and density significantly increased in

cases with nodal and distant metastasis. In their study, the frequency of distant metastasis increased with an increase in the microvessel count. However, other studies showed conflicting results. Van Hoef and colleagues [58] reported considerable variability in the microvessel count in different parts of the same tumor and between the readings of two evaluators. These investigators found no significant correlation between microvessel count and other tumor factors of prognostic value, and found no significant correlation between the microvessel count and DFS. Until these issues are resolved, microvessel count should not be used routinely for making treatment decisions in breast cancer patients.

Angiogenic growth factors have been identified that may have important prognostic utility. These include the vascular endothelial growth factor, the platelet-derived endothelial cell growth factor (also known as thymidine phosphorylase), and the fibroblast growth factor family [59].

Apoptosis-related prognostic markers

Programmed cell death, also known as apoptosis, is an endogenous cellular process whereby an external signal activates a metabolic pathway that results in cell death (Table 3) [60]. This form of cell death is commonly seen in breast cancer tissue. Apoptotic cells can be quantitated by light microscopy, and an apoptotic index can be calculated. However, the prognostic significance of the apoptotic index is not well defined. Wu and colleagues [61] reported a correlation between a low apoptotic index and decreased patient survival. However, other studies found no correlation between apoptotic index and prognosis [62–64].

Bcl-2 is a mitochondrial protein known to inhibit apoptosis triggered by chemotherapy and radiation therapy. Lower levels of apoptosis could lead to malignant cell accumulation and therefore to a more aggressive clinical course for the disease. Although Bcl-2 can block apoptosis *in vitro*, several studies have shown that Bcl-2 overexpression is associated with improved DFS rates [65]. This may be in part because of the close association between Bcl-2 expression and ER expression. Perhaps more important is the potential association between Bcl-2 expression and response to chemotherapy. Several studies have shown that patients with Bcl-2-negative breast cancer were more likely to respond to chemotherapy than patients with Bcl-2-positive tumors [66–68]. However, other studies found no association between Bcl-2 expression and the response to chemotherapy [69,70]. Further studies are needed to establish the role of Bcl-2 as a predictive factor of response to therapy.

Genomics

In addition to the markers already discussed, literally hundreds of other molecules have been evaluated as

Table 3

Members of the Bcl-2 family of apoptosis regulators

Inhibitor of apoptosis	Promoter of apoptosis
BCL-2	BAX
BCL-X _L	BAK
MCL-1	BOK
A-1/BFL-1	BAD
BCL-W	BID
BOO/DIVA	BIK
NR-13	BLK
	HRK
	BIM
	BNIP3
	NIX
	NOXA

potential prognostic factors. Breast cancer is a complex heterogeneous disease, and therefore evaluation of a handful of genes and/or proteins provides only limited prognostic information. High-throughput gene expression profiling using microarray technology is a promising new technology that has been applied to the classification of breast cancers [71–73], to prognosis [74–77], and to prediction of response to treatment [78].

Using cDNA microarrays, Perou and colleagues [71] classified invasive breast carcinomas into five subtypes based on their distinct gene expression profile (Norway/Stanford dataset). These included a luminal epithelial cell phenotype (subtypes A and B), a basal epithelial cell type phenotype, a HER-2 (+) phenotype, and a group of cancers expressing a 'normal-like' gene profile. Sorlie and colleagues [79] showed that patients whose tumors exhibited the basal-like and HER-2-positive subtypes had the worst survival rates, while the luminal epithelial type was associated with improved survival rates. Although initially the luminal subtype correlated with ER positivity, Sorlie and colleagues noted that the ER levels were not uniform among tumors classified as luminal or basal types.

van't Veer and colleagues [74] used a different microarray platform and identified a 'poor prognosis signature' that included 70 genes involved in regulation of the cell cycle, in invasion, in metastasis, and in angiogenesis. The 70-gene prognostic profile was validated by the same investigators in 295 consecutive patients with primary breast cancer [75]. Among the 295 patients, 180 had a poor-prognosis signature and 115 had a good-prognosis signature; the mean overall 10-year survival rates were

54.6% and 94.5%, respectively. At 10 years, the probability of remaining free of distant metastases was 50.6% in the group with a poor-prognosis signature and was 85.2% in the group with a good-prognosis signature. The estimated hazard ratio for distant metastases in the group with a poor-prognosis signature as compared with the group with the good-prognosis signature was 5.1 (95% confidence interval, 2.9–9.0; $P < 0.001$). This ratio remained significant when the groups were analyzed according to their lymph-node status. This prognostic signature had a strong independent value on multivariate analysis. Ongoing studies are validating these results in commercially available microarrays for potential clinical and diagnostic applications.

Sorlie and colleagues [80] reanalyzed their Norway/Stanford dataset, including 84 tissue samples from their previously published work [71,79] and 38 additional tumor samples from patients with locally advanced breast cancer treated with preoperative chemotherapy. The first gene list and the list used for the reanalyzed report had approximately 200 genes in common, and tumors could be classified in the five main gene clusters as previously described. In addition, Sorlie and colleagues attempted to validate their findings in two independent datasets reported by Van't Veer and colleagues [75] and by West and colleagues [81]. Ninety-seven tumors from the van't Veer and colleagues' study could be classified into the five subtypes, and these different breast cancer types were associated with prognosis. Patients with the luminal-A subtype had the best survival rates, while the worst survival rates were associated with the basal and HER-2 subtypes. However, van't Veer and colleagues based their analysis on 461 genes (out of 24,480). The dataset from West and colleagues, generated on an Affymetrix platform, could also be classified into the previously described subtypes after selecting 242 genes out of a total of 7129 genes.

One of the main shortcomings of microarray technology is the lack of validation of gene sets across platforms. For example, when Sorlie and colleagues [80] tested the prognostic impact of the 231 markers published by van't Veer and colleagues on the Norwegian cohort, the positive predictive value for DFS was only 47%. This may in part be due to the different patient cohorts and treatments. In fact, the differences in outcomes across studies are based on the subset of genes that was analyzed in all the studies, and the number of genes held in common across studies is limited.

Clinical trials are evaluating the prognostic and predictive value of gene expression profiles in patients with early-stage breast cancer. Chang and colleagues [82] evaluated gene expression profiles in tumors from 24 patients undergoing neoadjuvant docetaxel chemotherapy. Core

biopsies were obtained prior to initiation of chemotherapy, and cDNA analysis of RNA extracted from biopsy samples was completed using the HgU95-Av2 GeneChip (Affymetrix, Santa Clara, CA, USA). Differential patterns of expression of 92 genes correlated with docetaxel response ($P = 0.001$). Symmans and colleagues [83] showed that fine-needle aspiration yielded sufficient RNA for gene expression profiling. Since most breast cancer in patients is diagnosed at an early stage, the fine-needle aspiration approach may become acknowledged as the optimal way to obtain tissue for gene profiling. Pusztai and colleagues [78] extracted RNA from fine-needle aspiration specimens and identified a group of genes that predicted pathologic complete response to neoadjuvant chemotherapy.

Transcriptional profiling could until recently only be completed using fresh or frozen tissue, not using tissue from paraffin blocks. To overcome this limitation, several groups are developing methods to extract RNA from formalin-fixed, paraffin-embedded tissue for genomics studies. Ma and colleagues [84] microdissected breast cancer cells from paraffin-embedded tumors and measured expression on more than 20,000 genes in cancer cells using an Affymetrix platform. The authors were able to correlate gene expression signatures with prognosis. This is a step forward, and these findings should be validated in groups of patients treated homogeneously or not treated with adjuvant systemic therapy at all.

Several groups are evaluating the prognostic and predictive value of a multigene RT-PCR assay using paraffin-embedded tissue (Oncotype DX™; Genomic Health, Redwood City, CA, USA). Sixteen genes had significant prognostic value in three preliminary studies that included patients with early-stage breast cancer treated with adjuvant tamoxifen and/or chemotherapy [85–87]. Five genes were added as reference genes, and a recurrence score was developed. Paik and colleagues [85] showed that the multigene RT-PCR assay had a strong predictive value in patients with a history of node-negative, ER-positive tumors treated with tamoxifen in the adjuvant setting. The 10-year distant recurrence rate was 6.8% for patients with a low recurrence score, was 14.3% for patients with an intermediate recurrence score, and was 30.5% for patients with a high recurrence score.

A smaller study conducted at MD Anderson Cancer Center showed no relationship between OncotypeDX's recurrence score and distant recurrence-free survival in patients with node-negative breast cancer who had not received any adjuvant systemic therapy [88]. Although there may be many explanations for this finding, it is also possible that the model is good at predicting response or lack of response to tamoxifen but has limited prognostic power. More studies are needed to establish the prognostic role of this assay in clinical management.

Proteomics

In the postgenome era, scientists have turned to proteomics to understand complex biological systems. Proteomics is defined as the identification, characterization, and quantification of all proteins involved in a particular tissue, organ, or organism to provide accurate and comprehensive data about that system.

One of the methods most commonly used to study differences in protein expression between two samples (e.g. cancer and normal tissue) is two-dimensional gel electrophoresis. Highly sensitive mass spectrometry methods are currently being used together to identify greater numbers of lower abundance proteins that are differentially expressed in defined cell populations. Matrix-assisted laser desorption/ionization time-of-flight and surface-enhanced laser desorption/ionization time-of-flight analyses enable high-throughput characterization of lysates from even a very few tumor cells, and they may be best suited for clinical biomarker studies [89,90].

Novel technologies still in developmental phases will enable identification of validated targets in small biopsy specimens, including high-density protein, antibody, and lysate arrays [91,92]. No proteomics-based assay for assessing prognosis in breast cancer patients has yet been developed.

Conclusion

Prognostic and predictive molecular markers commonly used in clinical practice include Ki-67, ER, PR, and HER-2. From the National Institute of Health overview it was clear that, once basic pathology had been excluded, there was very little else that had been appropriately validated and in which there was good quality control. This issue of quality control is one of the most important challenges for validation of most molecular markers discussed.

Prognostic indices that integrate clinical, histologic, and molecular parameters will need to be developed and validated in conjunction with novel bioinformatic methodologies (i.e. artificial intelligence) to aid clinical decision-making. High-throughput cDNA microarray technologies and tumor array technologies are allowing the expression of literally thousands of genes and proteins to be analyzed at one time. Validation of these technologies in adequately powered prospective clinical trials will allow the integration of multiple molecular factors in the risk assessment and management of individual patients with breast cancer.

Competing interests

None declared.

Acknowledgements

This work was supported in part by the Nellie B Connally Breast Cancer Research Fund, and by grants from the National Cancer Institute (CA82119) and the Department of Defense (DAMD17-03-1-0429). The authors thank Rachel E Williams for editorial assistance.

References

1. Early Breast Cancer Trialists' Collaborative Group: **Systemic treatment of early breast cancer by hormonal, cytotoxic, or immune therapy. 133 randomised trials involving 31,000 recurrences and 24,000 deaths among 75,000 women.** *Lancet* 1992, **339**:1-15.
2. Early Breast Cancer Trialists' Collaborative Group: **Tamoxifen for early breast cancer: an overview of the randomised trials.** *Lancet* 1998, **351**:1451-1467.
3. Eifel P, Axelson JA, Costa J, Crowley J, Curran WJJ, Deshler A, Fulton S, Hendricks CB, Kemeny M, Kornblith AB, Louis TA, Markman M, Mayer R, Roter D: **National Institutes of Health Consensus Development Conference statement: adjuvant therapy for breast cancer, November 1-3, 2000.** *J Natl Cancer Inst* 2001, **93**:979-989.
4. Clark GM, Dressler LG, Owens MA, Pounds G, Oldaker T, McGuire WL: **Prediction of relapse or survival in patients with node-negative breast cancer by DNA flow cytometry.** *N Engl J Med* 1989, **320**:627-633.
5. Sahin AA, Ro J, Ro JY, Blick MB, el-Naggar AK, Ordonez NG, Fritsche HA, Smith TL, Hortobagyi GN, Ayala AG: **Ki-67 immunostaining in node-negative stage I/II breast carcinoma. Significant correlation with prognosis.** *Cancer* 1991, **68**:549-557.
6. Keshgegian AA, Cnaan A: **Proliferation markers in breast carcinoma. Mitotic figure count, S-phase fraction, proliferating cell nuclear antigen, Ki-67 and MIB-1.** *Am J Clin Pathol* 1995, **104**:42-49.
7. Veronese SM, Gambacorta M, Gottardi O, Scanzini F, Ferrari M, Lampertico P: **Proliferation index as a prognostic marker in breast cancer.** *Cancer* 1993, **71**:3926-3931.
8. Zhu X, Mancini MA, Chang KH, Liu CY, Chen CF, Shan B, Jones D, Yang-Feng TL, Lee WH: **Characterization of a novel 350-kilodalton nuclear phosphoprotein that is specifically involved in mitotic-phase progression.** *Mol Cell Biol* 1995, **15**:5017-5029.
9. Clark GM, Allred DC, Hilsenbeck SG, Chamness GC, Osborne CK, Jones D, Lee WH: **Mitotin (a new proliferation marker) correlates with clinical outcome in node-negative breast cancer.** *Cancer Res* 1997, **57**:5505-5508.
10. Sommer S, Fuqua SA: **Estrogen receptor and breast cancer.** *Semin Cancer Biol* 2001, **11**:339-352.
11. Kuiper GG, Enmark E, Peltö-Huikko M, Nilsson S, Gustafsson JA: **Cloning of a novel receptor expressed in rat prostate and ovary.** *Proc Natl Acad Sci USA* 1996, **93**:5925-5930.
12. Speirs V, Kerin MJ: **Prognostic significance of oestrogen receptor beta in breast cancer.** *Br J Surg* 2000, **87**:405-409.
13. Dotzlaw H, Leygue E, Watson PH, Murphy LC: **Estrogen receptor-beta messenger RNA expression in human breast tumor biopsies: relationship to steroid receptor status and regulation by progestins.** *Cancer Res* 1999, **59**:529-532.
14. Fuqua SA, Schiff R, Parra I, Friedrichs WE, Su JL, McKee DD, Slentz-Kesler K, Moore LB, Willson TM, Moore JT: **Expression of wild-type estrogen receptor beta and variant isoforms in human breast cancer.** *Cancer Res* 1999, **59**:5425-5428.
15. Su JL, McKee DD, Ellis B, Kadwell SH, Wisely GB, Moore LB, Triantafyllou JA, Kost TA, Fuqua S, Moore JT: **Production and characterization of an estrogen receptor beta subtype-specific mouse monoclonal antibody.** *Hybridoma* 2000, **19**:481-487.
16. Holmes FA, Fritsche HA, Loewy JW, Geitner AM, Sutton RC, Buzdar AU, Hortobagyi GN: **Measurement of estrogen and progesterone receptors in human breast tumors: enzyme immunoassay versus binding assay.** *J Clin Oncol* 1990, **8**:1025-1035.
17. Donegan WL: **Tumor-related prognostic factors for breast cancer.** *CA Cancer J Clin* 1997, **47**:28-51.
18. Shek LL, Godolphin W: **Survival with breast cancer: the importance of estrogen receptor quantity.** *Eur J Cancer Clin Oncol* 1989, **25**:243-250.
19. Hahnel R, Woodings T, Vivian AB: **Prognostic value of estrogen receptors in primary breast cancer.** *Cancer* 1979, **44**:671-675.
20. Thorpe SM, Rose C, Rasmussen BB, King WJ, DeSombre ER, Blough RM, Mouridsen HT, Rossing N, Andersen KW: **Steroid hormone receptors as prognostic indicators in primary breast cancer.** *Breast Cancer Res Treat* 1986, **7**:91-98.
21. Crowe JP, Jr, Gordon NH, Hubay CA, Shenk RR, Zollinger RM, Brumberg DJ, McGuire WL, Shuck JM: **Estrogen receptor deter-**

- mination and long term survival of patients with carcinoma of the breast. *Surg Gynecol Obstet* 1991, **173**:273-278.
22. McGuire WL, Tandon AK, Allred DC, Chamness GC, Clark GM: **How to use prognostic factors in axillary node-negative breast cancer patients.** *J Natl Cancer Inst* 1990, **82**:1006-1015.
 23. Fisher B, Fisher ER, Redmond C, Brown A: **Tumor nuclear grade, estrogen receptor, and progesterone receptor: their value alone or in combination as indicators of outcome following adjuvant therapy for breast cancer.** *Breast Cancer Res Treat* 1986, **7**:147-160.
 24. Allred DC, Bryant J, Land S, Paik S, Fisher E, Julian T, Margolese R, Smith R, Mamounas T, Osborne CK, Fisher B, Wolmark N: **Estrogen receptor expression as a predictive marker of the effectiveness of tamoxifen in the treatment of DCIS: findings from NSABP Protocol B-24 [abstract].** *Breast Cancer Res Treat* 2002, **76**:A36.
 25. Fuqua SA, Schiff R, Parra I, Moore JT, Mohsin SK, Osborne CK, Clark GM, Allred DC: **Estrogen receptor beta protein in human breast cancer: correlation with clinical tumor parameters.** *Cancer Res* 2003, **63**:2434-2439.
 26. Dickson RB, Lippman ME: **Growth factors in breast cancer.** *Endocr Rev* 1995, **16**:559-589.
 27. Bacus SS, Gudkov AV, Esteva FJ, Yarden Y: **Expression of erbB receptors and their ligands in breast cancer: implications to biological behavior and therapeutic response.** *Breast Dis* 2000, **11**:63-75.
 28. Di Fiore PP, Pierce JH, Kraus MH, Segatto O, King CR, Aaronson SA: **ErbB-2 is a potent oncogene when overexpressed in NIH/3T3 cells.** *Science* 1987, **237**:178-182.
 29. Guy CT, Webster MA, Schaller M, Parsons TJ, Cardiff RD, Muller WJ: **Expression of the neu protooncogene in the mammary epithelium of transgenic mice induces metastatic disease.** *Proc Natl Acad Sci USA* 1992, **89**:10578-10582.
 30. Esteva-Lorenzo FJ, Sastry L, King CR: **The erbB-2 gene: from research to application.** In *Hormones and Growth Factors in Development and Neoplasia*. Edited by Dickson RB, Salomon DS. New York: John Wiley & Sons; 1998:421-444.
 31. King CR, Kraus MH, Williams LT, Merlino GT, Pastan IH, Aaronson SA: **Human tumor cell lines with EGF receptor gene amplification in the absence of aberrant sized mRNAs.** *Nucleic Acids Res* 1985, **13**:8477-8486.
 32. Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL: **Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene.** *Science* 1987, **235**:177-182.
 33. Esteva FJ, Pusztai L, Symmans WF, Sneige N, Hortobagyi GN: **Clinical relevance of HER-2 amplification and overexpression in human cancers.** *Ref Gynecol Obst* 2000, **7**:267-276.
 34. Borg A, Tandon AK, Sigurdsson H, Clark GM, Ferno M, Fuqua SA, Killander D, McGuire WL: **HER-2/neu amplification predicts poor survival in node-positive breast cancer.** *Cancer Res* 1990, **50**:4332-4337.
 35. Allred DC, Clark GM, Tandon AK, Molina R, Tormey DC, Osborne CK, Gilchrist KW, Mansour EG, Abeloff M, Eudey L, et al.: **Her-2/neu in node-negative breast cancer: prognostic significance of overexpression influenced by the presence of in situ carcinoma.** *J Clin Oncol* 1992, **10**:599-605.
 36. Wood WC, Budman DR, Korzun AH, Cooper MR, Younger J, Hart RD, Moore A, Ellerton JA, Norton L, Ferree CR: **Dose and dose intensity of adjuvant chemotherapy for stage II, node-positive breast carcinoma.** *N Engl J Med* 1994, **330**:1253-1259.
 37. Muss HB, Thor AD, Berry DA, Kute T, Liu ET, Koerner F, Cirrincione CT, Budman DR, Wood WC, Barcos M, Henderson IC: **c-erbB-2 expression and response to adjuvant therapy in women with node-positive early breast cancer.** *N Engl J Med* 1994, **330**:1260-1266.
 38. Thor AD, Berry DA, Budman DR, Muss HB, Kute T, Henderson IC, Barcos M, Cirrincione C, Edgerton S, Allred C, Norton L, Liu ET: **erbB-2, p53, and efficacy of adjuvant therapy in lymph node-positive breast cancer.** *J Natl Cancer Inst* 1998, **90**:1346-1360.
 39. Paik S, Bryant J, Park C, Fisher B, Tan-Chiu E, Hyams D, Fisher ER, Lippman ME, Wickerham DL, Wolmark N: **erbB-2 and response to doxorubicin in patients with axillary lymph node-positive, hormone receptor-negative breast cancer.** *J Natl Cancer Inst* 1998, **90**:1361-1370.
 40. Ravdin PM, Green S, Albain KS, Boucher V, Ingle J, Pritchard K, Shepard L, Davidson N, Hayes DF, Clark GM, Martino S, Osborne CK, Allred DC: **Initial report of the SWOG biological correlative study of C-erbB-2 expression as a predictor of outcome in a trial comparing adjuvant CAF T with tamoxifen (T) alone [abstract].** *Proc Am Soc Clin Oncol* 1998, **17**:A374.
 41. Van Poznak C, Tan L, Panageas KS, Arroyo CD, Hudis C, Norton L, Seidman AD: **Assessment of molecular markers of clinical sensitivity to single-agent taxane therapy for metastatic breast cancer.** *J Clin Oncol* 2002, **20**:2319-2326.
 42. De Placido S, De Laurentiis M, Carlomagno C, Gallo C, Perrone F, Pepe S, Ruggiero A, Marinelli A, Pagliarulo C, Panico L, Pettinato G, Petrella G, Bianco AR: **Twenty-year results of the Naples GUN randomized trial: Predictive factors of adjuvant tamoxifen efficacy in early breast cancer.** *Clin Cancer Res* 2003, **9**:1039-1046.
 43. Elledge RM, Green S, Ciocca D, Pugh R, Allred DC, Clark GM, Hill J, Ravdin P, O'Sullivan J, Martino S, Osborne CK: **HER-2 expression and response to tamoxifen in estrogen receptor-positive breast cancer: a Southwest Oncology Group Study.** *Clin Cancer Res* 1998, **4**:7-12.
 44. Osborne CK, Bardou V, Hopp TA, Chamness GC, Hilsenbeck SG, Fuqua SA, Wong J, Allred DC, Clark GM, Schiff R: **Role of the estrogen receptor coactivator AIB1 (SRC-3) and HER-2/neu in tamoxifen resistance in breast cancer.** *J Natl Cancer Inst* 2003, **95**:353-361.
 45. Press M, Anderson S, Dybdal N, Lieberman G, Mass R: **Fluorescence in situ hybridization (FISH) is superior to immunohistochemistry (IHC) for determining HER2 status: the Herceptin (R) experience [abstract].** *Mod Pathol* 2002, **15**:A47.
 46. Seidman AD, Fornier M, Esteva FJ, Tan L, Kaptain S, Bach A, Panageas KS, Arroyo C, Valero V, Currie V, Gilewski T, Theodoulou M, Moynahan ME, Moasser M, Sklarin N, Dickler M, D'Andrea G, Cristofanilli M, Rivera E, Hortobagyi GN, Norton L, Hudis C: **Weekly trastuzumab and paclitaxel therapy for metastatic breast cancer with analysis of efficacy by HER2 immunophenotype and gene amplification.** *J Clin Oncol* 2001, **19**:2587-2595.
 47. Janicke F, Schmitt M, Pache L, Ulm K, Harbeck N, Hofler H, Graeff H: **Urokinase (uPA) and its inhibitor PAI-1 are strong and independent prognostic factors in node-negative breast cancer.** *Breast Cancer Res Treat* 1993, **24**:195-208.
 48. Bouchet C, Spyrtos F, Martin PM, Hacene K, Gentile A, Oglobine J: **Prognostic value of urokinase-type plasminogen activator (uPA) and plasminogen activator inhibitors PAI-1 and PAI-2 in breast carcinomas.** *Br J Cancer* 1994, **69**:398-405.
 49. Grondahl-Hansen J, Hilsenbeck SG, Christensen IJ, Clark GM, Osborne CK, Brunner N: **Prognostic significance of PAI-1 and uPA in cytosolic extracts obtained from node-positive breast cancer patients.** *Breast Cancer Res Treat* 1997, **43**:153-163.
 50. Broet P, Spyrtos F, Romain S, Quillien V, Daver A, Ricolleau G, Rallet A, Toulas C, Asselain B: **Prognostic value of uPA and p53 accumulation measured by quantitative biochemical assays in 1245 primary breast cancer patients: a multicenter study.** *Br J Cancer* 1999, **80**:536-545.
 51. de Witte JH, Sweep CG, Klijn JG, Grebenschikov N, Peters HA, Look MP, van Tienoven TH, Heuvel JJ, van Putten WL, Benraad TJ, Foekens JA: **Prognostic impact of urokinase-type plasminogen activator (uPA) and its inhibitor (PAI-1) in cytosols and pellet extracts derived from 892 breast cancer patients.** *Br J Cancer* 1999, **79**:1190-1198.
 52. Janicke F, Prechtel A, Thomssen C, Harbeck N, Meisner C, Untch M, Sweep CGJF, Selbmann HK, Graeff H, Schmitt M: **Randomized adjuvant chemotherapy trial in high-risk, lymph node-negative breast cancer patients identified by urokinase-type plasminogen activator and plasminogen activator inhibitor type 1.** *J Natl Cancer Inst* 2001, **93**:913-920.
 53. Harbeck N, Meisner C, Prechtel A, Untch M, Selbmann HK, Sweep F, Graeff H, Schmitt M, Jaenicke F, Thomssen C: **Level-I evidence for prognostic and predictive impact of uPA and PAI-1 in node-negative breast cancer provided by second scheduled analysis of multicenter Chemo-N-0 therapy trial [abstract].** *Breast Cancer Res Treat* 2001, **89**:A213.
 54. Zemzoum I, Kates RE, Ross JS, Detmar P, Dutta M, Henrichs C, Yurdseven S, Hofler H, Kiechle M, Schmitt M, Harbeck N: **Invasion factors uPA/PAI-1 and HER2 status provide independent and complementary information on patient outcome in node-negative breast cancer.** *J Clin Oncol* 2003, **21**:1022-1028.

55. Harbeck N, Schmitt M, Kates RE, Kiechle M, Zemzoum I, Janicke F, Thomssen C: **Clinical utility of urokinase-type plasminogen activator and plasminogen activator inhibitor-1 determination in primary breast cancer tissue for individualized therapy concepts.** *Clin Breast Cancer* 2002, **3**:196-200.
56. Folkman J: **Angiogenesis in cancer, vascular, rheumatoid and other disease.** *Nat Med* 1995, **1**:27-31.
57. Weidner N, Semple JP, Welch WR, Folkman J: **Tumor angiogenesis and metastasis—correlation in invasive breast carcinoma.** *N Engl J Med* 1991, **324**:1-8.
58. Van Hoef ME, Knox WF, Dhesi SS, Howell A, Schor AM: **Assessment of tumour vascularity as a prognostic factor in lymph node negative invasive breast cancer.** *Eur J Cancer* 1993, **29A**: 1141-1145.
59. Gasparini G: **Clinical significance of determination of surrogate markers of angiogenesis in breast cancer.** *Crit Rev Oncol Hematol* 2001, **37**:97-114.
60. Wyllie AH, Kerr JF, Currie AR: **Cell death: the significance of apoptosis.** *Int Rev Cytol* 1980, **68**:251-306.
61. Wu J, Shen ZZ, Lu JS, Jiang M, Han QX, Fontana JA, Barsky SH, Shao ZM: **Prognostic role of p27Kip1 and apoptosis in human breast cancer.** *Br J Cancer* 1999, **79**:1572-1578.
62. Lipponen P, Aaltomaa S, Kosma VM, Syrjanen K: **Apoptosis in breast cancer as related to histopathological characteristics and prognosis.** *Eur J Cancer* 1994, **30A**:2068-2073.
63. Zhang GJ, Kimijima I, Abe R, Watanabe T, Kanno M, Hara K, Tsuchiya A: **Apoptotic index correlates to bcl-2 and p53 protein expression, histological grade and prognosis in invasive breast cancers.** *Anticancer Res* 1998, **18**:1989-1998.
64. Liu SQ, Edgerton SM, Moore DHI, Thor AD: **Measures of cell turnover (proliferation and apoptosis) and their association with survival in breast cancer.** *Clin Cancer Res* 2001, **7**:1716-1723.
65. Gasparini G, Barbareschi M, Doglioni C, Palma PD, Mauri FA, Boracchi P, Bevilacqua P, Caffo O, Morelli L, Verderio P: **Expression of bcl-2 protein predicts efficacy of adjuvant treatments in operable node-positive breast cancer.** *Clin Cancer Res* 1995, **1**:189-198.
66. Bonetti A, Zaninelli M, Leone R, Cetto GL, Pelosi G, Biolo S, Menghi A, Manfrin E, Bonetti F, Piubello Q: **bcl-2 but not p53 expression is associated with resistance to chemotherapy in advanced breast cancer.** *Clin Cancer Res* 1998, **4**:2331-2336.
67. Pusztai L, Krishnamurthy S, Perez-Cardona J, Sneige N, Esteva FJ, Volchenok M, Breitenfelder P, Kau SW, Takayama S, Krajewski S, Reed JC, Hortobagyi GN, Bast RC: **Expression of BAG-1 and BCL-2 proteins before and after neoadjuvant chemotherapy of locally advanced breast cancer.** *Cancer Invest* 2004, in press.
68. Buchholz TA, Davis DW, McConkey DJ, Symmans WF, Valero V, Jhingran A, Tucker SL, Pusztai L, Cristofanilli M, Esteva FJ, Hortobagyi GN, Sahin AA: **Chemotherapy-induced apoptosis and Bcl-2 levels correlate with breast cancer response to chemotherapy.** *Cancer J* 2003, **9**:33-41.
69. Bottini A, Berruti A, Bersiga A, Brizzi MP, Brunelli A, Gorzegno G, DiMarco B, Aguggini S, Bolsi G, Cirillo F, Filippini L, Betri E, Bertoli G, Alquati P, Dogliotti L: **p53 but not bcl-2 immunostaining is predictive of poor clinical complete response to primary chemotherapy in breast cancer patients.** *Clin Cancer Res* 2000, **6**:2751-2758.
70. Poelman SM, Adeyanju MO, Robertson MA, Recant WM, Karrison T, Fleming GF, Olopade OI, Conzen SD: **Human breast cancer susceptibility to paclitaxel therapy is independent of Bcl-2 expression.** *Clin Cancer Res* 2000, **6**:4043-4048.
71. Perou CM, Sorlie T, Eisen MB, van de RM, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, Fluge O, Pergamenschikov A, Williams C, Zhu SX, Lonning PE, Borresen-Dale AL, Brown PO, Botstein D: **Molecular portraits of human breast tumours.** *Nature* 2000, **406**:747-752.
72. Pusztai L, Ayers M, Stec J, Clark E, Hess K, Stivers D, Damokosh A, Sneige N, Buchholz TA, Esteva FJ, Arun B, Cristofanilli M, Booser D, Rosales M, Valero V, Adams C, Hortobagyi GN, Symmans WF: **Gene expression profiles obtained from fine-needle aspirations of breast cancer reliably identify routine prognostic markers and reveal large-scale molecular differences between estrogen-negative and estrogen-positive tumors.** *Clin Cancer Res* 2003, **9**:2406-2415.
73. Pusztai L, Sotiriou C, Buchholz TA, Meric F, Symmans WF, Esteva FJ, Sahin A, Liu ET, Hortobagyi GN: **Molecular profiles of invasive mucinous and ductal carcinomas of the breast: a molecular case study.** *Cancer Genet Cytogenet* 2003, **141**:148-153.
74. van't Veer LJ, Dai H, van de Vijver MJ, He YD, Hart AA, Mao M, Peterse HL, van der KK, Marton MJ, Witteveen AT, Schreiber GJ, Kerkhoven RM, Roberts C, Linsley PS, Bernards R, Friend SH: **Gene expression profiling predicts clinical outcome of breast cancer.** *Nature* 2002, **415**:530-536.
75. van de Vijver MJ, He YD, van't Veer LJ, Dai H, Hart AA, Voskuil DW, Schreiber GJ, Peterse JL, Roberts C, Marton MJ, Parrish M, Atsma D, Witteveen A, Glas A, Delahaye L, van der Velde, Bartelink H, Rodenhuis S, Rutgers ET, Friend SH, Bernards R: **A gene-expression signature as a predictor of survival in breast cancer.** *N Engl J Med* 2002, **347**:1999-2009.
76. Bertucci F, Houlgatte R, Granjeaud S, Nasser V, Loriod B, Beau-doing E, Hingamp P, Jacquemier J, Viens P, Birnbaum D, Nguyen C: **Prognosis of breast cancer and gene expression profiling using DNA arrays.** *Ann NY Acad Sci* 2002, **975**:217-231.
77. Bertucci F, Nasser V, Granjeaud S, Eisinger F, Adelaide J, Tagett R, Loriod A, Giaconia A, Benziane A, Devillard E, Jacquemier J, Viens P, Nguyen C, Birnbaum D, Houlgatte M: **Gene expression profiles of poor-prognosis primary breast cancer correlate with survival.** *Hum Mol Genet* 2002, **11**:863-872.
78. Pusztai L, Ayers M, Symmans WF, Damokosh A, Hess K, Valero V, Clark E, Ross J, Hortobagyi GN, Stec J: **Emerging science: prospective validation of gene expression profiling-based prediction of complete pathologic response to neoadjuvant paclitaxel/FAC chemotherapy in breast cancer [abstract].** *Proc Am Soc Clin Oncol* 2003, **22**:A1.
79. Sorlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, Hastie T, Eisen MB, van de Rijn M, Jeffrey SS, Thorsen T, Quist H, Matese JC, Brown PO, Botstein D, Lonning PE, Borresen-Dale AL: **Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications.** *Proc Natl Acad Sci USA* 2001, **98**:10869-10874.
80. Sorlie T, Tibshirani R, Parker J, Hastie T, Marron JS, Nobel A, Deng S, Johnsen H, Pesich R, Geisler S, Demeter J, Perou CM, Lonning PE, Brown PO, Borresen-Dale AL, Botstein D: **Repeated observation of breast tumor subtypes in independent gene expression data sets.** *Proc Natl Acad Sci USA* 2003, **100**: 8418-8423.
81. West M, Blanchette C, Dressman H, Huang E, Ishida S, Spang R, Zuzan H, Olson JA, Jr, Marks JR, Nevins JR: **Predicting the clinical status of human breast cancer by using gene expression profiles.** *Proc Natl Acad Sci USA* 2001, **98**:11462-11467.
82. Chang JC, Wooten EC, Tsimelzon A, Hilsenbeck SG, Gutierrez MC, Elledge R, Mohsin S, Osborne CK, Chamness GC, Allred DC, O'Connell P: **Gene expression profiling for the prediction of therapeutic response to docetaxel in patients with breast cancer.** *Lancet* 2003, **362**:362-369.
83. Symmans WF, Ayers M, Clark EA, Stec J, Hess KR, Sneige N, Buchholz TA, Krishnamurthy S, Ibrahim NK, Buzdar AU, Theriault RL, Rosales MF, Thomas ES, Gwyn KM, Green MC, Syed AR, Hortobagyi GN, Pusztai L: **Total RNA yield and microarray gene expression profiles from fine-needle aspiration biopsy and core-needle biopsy samples of breast carcinoma.** *Cancer* 2003, **97**:2960-2971.
84. Ma X-J, Wang W, Salunga R, Tuggle T, Stecker K, Baer TM, Erlander MG, Witliff JL: **Gene expression signatures associated with clinical outcome in breast cancer via laser capture microdissection [abstract].** *Breast Cancer Res Treat* 2003, **82**: A29.
85. Paik S, Shak S, Tang G, Kim C, Baker J, Cronin M, Baehner R, Walker M, Watson D, Park T, Bryant J, Wolmark N: **Multi-gene RT-PCR assay for predicting recurrence in node negative breast cancer patients - NSABP studies B-20 and B-14 [abstract].** *Breast Cancer Res Treat* 2003, **82**:A16.
86. Cobleigh MA, Bitterman P, Baker J, Cronin M, Liu M-L, Borchik B, Tabesh B, Mosquera JM, Walker MG, Shak S: **Tumor gene expression predicts distant disease-free survival (DDFS) in breast cancer patients with 10 or more positive nodes: high throughput RT-PCR assay of paraffin-embedded tumor tissues [abstract].** *Proc Am Soc Clin Oncol* 2003, **22**:A3415.
87. Esteban J, Baker J, Cronin M, Liu M, Llamas MG, Walker MG, Mena R, Shak S: **Tumor gene expression and prognosis in breast cancer: multi-gene RT-PCR assay of paraffin-embedded tissue [abstract].** *Proc Am Soc Clin Oncol* 2003, **22**: A3416.

88. Esteva FJ, Sahin AA, Coombes K, Baker J, Cronin M, Walker M, Watson D, Cristofanilli M, Shak S, Hortobagyi GN: **Multi-gene RT-PCR assay for predicting recurrence in node negative breast cancer patients – M. D. Anderson Clinical Validation Study [abstract]**. *Breast Cancer Res Treat* 2003, **82**:A17.
89. Wulfkuhle JD, McLean KC, Paweletz CP, Sgroi DC, Trock BJ, Steeg PS, Petricoin EF: **New approaches to proteomic analysis of breast cancer**. *Proteomics* 2001, **1**:1205-1215.
90. Madoz-Gurpide J, Wang H, Misek DE, Brichory F, Hanash SM: **Protein based microarrays: a tool for probing the proteome of cancer cells and tissues**. *Proteomics* 2001, **1**:1279-1287.
91. Liotta LA, Espina V, Mehta AI, Calvert V, Rosenblatt K, Geho D, Munson PJ, Young L, Wulfkuhle J, Petricoin EF, III: **Protein microarrays: meeting analytical challenges for clinical applications**. *Cancer Cell* 2003, **3**:317-325.
92. Wulfkuhle JD, Liotta LA, Petricoin EF: **Proteomic applications for the early detection of cancer**. *Nat Rev Cancer* 2003, **3**:267-275.

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